COMPARATIVE PATHOLOGICAL STUDIES ON BACTERIAL AFFECTIONS OF LIVER IN FARM ANIMALS

Mohamed G.E. Borai, Abdel-Rehiem A. Nagi, Mahmoud S. Gab-Allah, Abdel-Baset I. El-Mashad and Shawky A. Moustafa.

Department of pathology, Faculty of Veterinary Medicine, Benha University

Corresponding author: e-mail: mohamedgouda2013@yahoo.com

Abstract

Out of the examined livers of some farm animals (1000 sheep, 825 cattle, 1740 buffaloes and 1360 camels) of different ages and sexes slaughtered at abattoirs in Cairo, Giza and Kalubia provinces, during the period from January 2011 to March 2013, the incidence of the bacterial affections was 5%, 3.03%, 1.29% and 1.91%, respectively. Bacteriological examination revealed that *Staphylococcus aureus, Corynebacterium pyogenes* and *Escherichia coli* were the most common aerobic bacteria while *Clostridium perfringens* and *Fusobacterium necrophorum* were the most common anaerobic bacteria isolated from the affected livers. *Corynebacterium ovis* was isolated from six cases of sheep. Histopathologically, suppurative hepatitis was the commonest lesion usually in the form of abscesses. Coagulative necrosis was noticed in cases where anaerobes were isolated.

KEY WORDS: Bacterial affections, Farm animals, *Staph. aureus, Coryne. pyogenes, E.coli,* Histopatholog, Hepatitis.

Introduction

Liver is considered to be the most important organ for mammalian metabolism thus any disturbance in this organ will reflect on the general health causing great economic losses in animal production [1]. Liver lesions are common and they indicate the presence of disease in other organs and systems as the liver acts as a catchment for the vast absorptive area of the gut, with all its resident microorganisms [2]. Liver abscesses in feedlot cattle, represent a major economic liability to producers, packers and ultimately consumers [3]. *Staphylococcus aureus, Corynebacterium pyogenes* and *Escherichia coli* were the predominant isolates from liver abscesses [4-7]. *Corynebacterium ovis* was also isolated from liver abscesses in sheep [8, 9]. *Clostridium species* was incriminated in liver affections by [10 -12]. *Fusobacterium necrophorum* is considered to be one of the most common causes of hepatic abscesses in ruminant [1, 13]. The present investigation was undertaken to study the major bacterial causes of liver lesions in different farm animals with comparative description of gross and

histopathological changes accompanied them.

Materials and Method

Animals: During the period extended from January 2011 to March 2013, livers of (1000 sheep, 825 cattle, 1740 buffaloes and 1360 camels) of different ages and sexes were randomly collected from the main abattoirs of Cairo, Giza and Kalubia provinces.

Sampling: Representative samples taken from affected livers were divided into two parts, one part was collected in plastic bags and transferred in an ice box to the laboratory for bacteriological examination, and the second one was immersed in 10% formalin for histopathological examination.

A)Bacteriological examination: The surface of the liver tissue was sterilized by a hot spatula, incised with a sterile scalpel and a sterilized platinum loop was inserted in tissue. A loopful of the liver content was inoculated into nutrient agar, 5% sheep blood agar and MacConkey agar and incubated aerobically at 37°C for 24-48 hours. Growing colonies were picked up and subcultured. The isolates were identified according to the colony morphology, pigment production, Gram's stain, as well as biochemical characters [14]. Another loopfuls were inoculated into two tubes of freshly prepared cooked meat broth, one of them was heated at 80°C for 10 minutes to eliminate non spore forming organisms while the other tube was left without heating, both tubes were incubated anaerobically at 37°C for 48 hours. A loopful from each heated tube was streaked onto blood agar plate for isolation of spore forming anaerobes while, loopfuls from each non heated tube were streaked onto neomycin blood agar and brain-heart infusion blood agar plates for isolation of *Clostridium* perfringens and non spore forming anaerobes, respectively. All plates were examined after anaerobic incubation for 2-3 days and each isolate was subcultured for identification according to [15].

B)Histopathological examination: specimens from affected livers were preserved in 10 %formalin .After proper fixation, the specimens were dehydrated in ascending grades of ethyl alcohol, then cleared in xylol and embedded in paraffin .Thin tissue sections about 5 microns in thickness were prepared and stained with hematoxylin and eosin stain for general microscopic examination according to [16].

Results

Table (1):	The	different	isolated	bacteria	among	the	total	affected	cases	of	livers
of sheep:											

Single isolates	No	%	Mixed isolates	No	%
Staphylococcus aureus	5	10	Staphylococcus aureus +	1	2
Corynebacterium pyogenes	10	20	Staphylococcus aureus + Pseudomonas aeruginosa	1	2
E. coli	7	14	Staphylococcus aureus + Proteus vulgaris	1	2
Corynebacterium ovis	4	8	Staphylococcus aureus + Klebseilla pneumonae	1	2
Pasturella haemolytica	1	2	Staphylococcus aureus + Peptostreptococcus anaerobious	2	4
Clostridium perfringens	6	12	Corynebacterium pyogenes + Corynebacterium ovis	1	2
Clostridium sordelli	1	2	Corynebacterium pyogenes +Fusobacterium necrophorum	1	2
Subtotal	34	68	Corynebacterium ovis + Clostridium perfringens	1	2
			E. coli + Clostridium perfringens + Clostridium sordelli	2	4
			E. coli + Clostridium perfringens	3	6
			Pasturella haemolytica +	1	2
			Fusobacterium necrophorum		
			Fusobacterium necrophorum +	1	2
			Peptostreptococcus anaerobious	1(22
	10	52			
	50	100			

Table (2): The different Isolated Bacteria among the total affected cases of livers of cattle:

Single isolate	No	%	Mixed isolates	No	%
Staphylococcus aureus	1	4	Staphylococcus aureus + Clostridium perfringens	2	8
Corynebacterium pyogenes	1	4	Staphylococcus aureus + Klebseilla pneumoniae	1	4
Fusobacterium necrophorum	1	4	Staphylococcus aureus + Fusobacterium necrophorum	1	4
Subtotal	3	12	Corynebacterium pyogenes + Fusobacterium necrophorum	6	24
			E- coli + Clostridium perfringens	5	20
			E. coli + Peptostreptococcus anaerobious	1	4
			Klebseilla pneumonae + Clostridium perfringens	1	4
			Proteus vulgaris + Clostridium perfringens	2	8
			Clostridium sporogenes + Clostridium perfringens	2	8
			Clostridium sordelli +	1	4
			Peptostreptococcus anaerobious		
			Subtotal	22	88
		Total		25	100

Single isolate	No	%	Mixed isolates	No	%
Staphylococcus aureus	2	10.53	Staphylococcus aureus +	3	15.79
			Fusobacterium necrophorum		
E. coli	2	10.53	Staphylococcus aureus +	1	5.26
			Clostridium sordelli		
Fusobacterium	2	10.53	Staphylococcus aureus +	1	5.26
necrophorum			Peptostreptococcus anaerobious		
Clostridium perfringens	2	10.53	Corynebacterium pyogenes+	3	15.79
			Fusobacterium necrophorum		
Subotal	8	42.11	E- coli + Fusobacterium	1	5.26
	-		necrophorum		
			E- coli + Clostridium perfringens	1	5.26
			E- coli + Streptococcus	1	5.26
			agalactiae		
			Subotal	11	57.89
		Total		19	100

Table (3): The different isolated bacteria among the total affected cases of livers of buffaloes:

Table (4): The different isolated bacteria among the total affected cases of livers of camels:

Single isolate	No	%	Mixed isolates	No	%
Staphylococcus aureus	3	11 54	Staphylococcus aureus +	1	3 85
Suprifice ceus aureus	5	11.01	Fusobacterium necrophorum		5.05
Corynebacterium pyogenes	5	19.23	Corynebacterium pyogenes	1	3.85
E coli	1	3 85	+Streptococcus pneumoniae	1	3 85
L. con	1	5.05	Fusobacterium necrophorum	1	5.65
Streptococcus pyogenes	1	3.85	Corynebacterium pyogenes +	1	3.85
Strantogoggus proumonio	1	2 95	Clostridium sporogenes	4	15 29
suepiococcus pneumonia	1	5.85	perfringens	4	15.56
Clostridium perfringens	1	3.85	E- coli + Fusobacterium	3	11.54
			necrophorum		
Subotal	12	46.15	E-coli+Peptostreptococcus anaerobious	1	3.85
			E- coli + Clostridium sordelli	1	3.85
			Streptococcus pyogenes +	1	3.85
			Clostridium perfringens		
			Subotal	14	53.85
	Total			26	100

<u>1-Staphylococcus aureus:</u>

The frequency related to the infected cases was 22 % (sheep), 20 % (cattle), 36.84 % (buffaloes) and 15.38 % (camels) isolated in single and mixed form as shown in tables (1-4). Macroscopically, affected livers showed areas of congestion and haemorrhages surrounding abscesses which contained whitish pus or cheesy material (fig.1). Microscopically, irregular areas of extravasated blood were seen (fig.2). Abscesses

formed from finely granular eosinophilic necrotic debris surrounded with leucocytes and C.T. capsule were obtained (fig.3). Microscopic abscesses represented by focal neutrophilic cellular infiltration were found in buffaloes and camels, meanwhile chronic calcified abscesses were detected in cases of sheep and camels (fig.4).

2- Corynebacterium pyogenes and Corynebacterium ovis:

The frequency of Coryne. pyogenes was 24% in sheep, 28% in cattle, 15.79 % in buffaloes and 30.77 % in camels meanwhile Coryne. ovis was recorded only in sheep in 12% (tables, 1-4). Macroscopically, multiple abscesses of a centimeter in diameter filled with greenish-white viscid pus were detected (fig. 5). Irregular greenish-white areas on the hepatic surface were seen in cattle while reddish and yellowish areas were seen in some cases of buffaloes and camels. Moreover, greyish white cirrhotic patches were noticed in some cases of camels. Microscopically, necrotic eosinophilic center surrounded with inflammatory cells and connective tissue capsule represented abscess formation (fig. 6). Area of dystrophic calcification was recorded in some cases of sheep and camels. Suppurative hepatitis represented by neutrophilic cellular infiltration in between the hepatic cells which showed areas of coagulative necrosis were noticed in cases of cattle (fig. 7). In cases infected with Coryne. pyogenes mixed with Fusobacterium necrophorum, hemorrhages and degenerative changes in the form of vacuolar and hydropic degeneration were seen in buffaloes and camels and in the form of fatty change together with the presence of coagulative necrosis in camel(fig. 8). Cirrhosis and mononuclear leucocytic infiltration were observed in some cases of camels (fig. 9).

<u>3- E. coli:</u>

The frequency was 24 % (sheep), 24 % (cattle), 26.32 % (buffaloes) and 38.46% (camels) in pure state and mixed mainly with anaerobes (tables, 1-4). Grossly, Abscesses up to one centimeter in diameter containing creamy viscid pus were found (fig.10). Microscopically, abscesses represented by necrotic areas infiltrated with neutrophils and surrounded with connective tissue capsule and mononuclear cells were obtained (fig. 11). Some abscesses showed chronicity with calcium deposition in some cases of cattle. Moreover, one case of camel's liver infected with *E. coli* showed polymorphonuclear and mononuclear leucocytes diffusely infiltrated and replaced necrotic hepatocytes. Other cases infected with *E. coli* mixed with anaerobes showed mononuclear leucocytic infiltration with the presence of fatty change of hepatocytes and congestion of the hepatic sinusoids, in addition to hyperplasia and desquamation of the biliary epithelium (fig. 12)



- **Fig. (1):** Liver of sheep infected with *Staph. aureus* showing multiple grayish white abscesses of 3 cm diameters containing whitish cheesy material and surrounded by reddish hyperemic zone or pale yellowish area on the diaphragmatic surface of the right lobe (arrow).
- **Fig. (2):** Liver of buffalo infected with *Staph. aureus* showing congestion of the hepatic blood vessels and sinusoids with individualization of the hepatocytes (H&E stain x300).



- Fig. (3): Liver of camel infected with *Staph. aureus* showing focal suppurative hepatitis characterized by finely granular eosinophilic and basophilic substance infiltrated with neutrophils and surrounded by fibrous capsule (H&E stain x800).
- Fig. (4): Liver of sheep infected with *Staph. aureus* showing old calcified abscesses which formed from dark purplish calcified areas surrounded with thick fibrous capsules (H&E stain x300).





- Fig. (5): Liver of sheep infected with *Coryne. pyogenes* showing multiple yellowish white different sizes abscesses 2 to 5 mm diameter scattered allover the hepatic tissue.
- Fig. (6): Liver of buffalo infected with *Coryne. pyogenes* showing abscess formed from caseous necrosis surrounded with zone of neutrophilic cellular infiltration followed by fibrous capsule infiltrated with leucocytes (H&E stain x300).



- **Fig. (7):** Liver of cattle infected with *Coryne. pyogenes* showing suppurative hepatitis manifested by heavy neutrophilic cellular infiltration replacing the hepatic cells (H&E stain x300).
- **Fig. (8):** Liver of camel infected with *Coryne. pyogenes* mixed with *F. necrophorum* showing diffuse hemorrhage and coagulative necrosis in the hepatic parenchyma (H&E stain x300).



- **Fig. (9):** Liver of camel infected with *Coryne. pyogenes* showing chronic hepatitis characterized by cirrhosis and mononuclear leucocytic cellular infiltration with hyperplasia of the epithelial cells of the bile ducts (H&E stain x300).
- **Fig.(10):**Liver of buffalo infected with *E. coli* showing whitish multiple circumscribed abscesses of different sizes on the visceral surface. On cutting, grayish white pus mixed with caseated flakes flew down (arrow).



- **Fig.(11):** Liver of sheep infected with *E. coli* showing focal suppurative inflammation surrounded by mononuclear and neutrophilic cellular infiltration and connective tissue capsule (H&E stain x300).
- **Fig. (12):** Liver of camel infected with *E. coli* showing diffuse leucocytic cellular infiltration of the hepatic tissue especially neutrophils and lymphocytes with mild degree of fatty change of the hepatocytes (H&E stain x600).

Discussion

Comparing to the prevalence of hepatic bacterial affections, it represented 4.4% of sheep slaughtered in Kalubia governorate [17], 2.92 % of cattle in Beni-Suef [18] and 2.5 % and 5.3 % of camels in Cairo and Sharkia governorates [10. 11], respectively, in the present study prevalence of liver lesion due to bacterial infection was 5 % in sheep, 3.03 % in cattle, 1.29 % in buffaloes and 1.91 % in camels. The difference of the frequencies of the hepatic bacterial affections between species of our study seemed to be due to the difference in management practices and nutrition of these species according to [9]. It should be noted that although many factors lead to bacterial gain to the liver tissue but two major causes were incriminated; the first is grain overload which causes steep decline in rumen pH and induces atony and damage of the ruminal wall. Ruminitis enhance some ruminal bacteria to reach the portal vein and finally causing hepatic abscess [19]. The other risk factor is migrating intestinal parasites which pre-optimize the conditions for a secondary bacterial infection [20]. Bacteriological examination in our study revealed that, the main bacterial isolates from the collected liver samples were Staph aureus (22 %, 20 %, 36.84%, 15.38%), Corynebacterium pyogenes (24%, 28%, 15.79%, 30.77%), E. coli (24%, 24%, 26.32 %, 38.46%), Fusobacterium necrophorum (6%, 32%, 47.37%, 19.23%), Clostridium perfringens (26%, 48%, 15.79 %, 23.08%), Clostridium sordellii (6%, 4%, 5.26%, 3.85%), Peptostreptococcus anerobius (6 %, 8%, 5.26%, 3.85%) and Mixed isolates (32%, 88%, 57.89, 53.85) in sheep, cattle, buffaloes and camels, respectively. These results were in a partial agreement with that mentioned by [17] who found that, the main bacterial isolates of sheep liver at Cairo and Kalubia were *Staphylococcus* (13.64%), Streptococcus (9.09%), E.coli (36.36%), Micrococcus (22.73%) and Pseudomonas (18.18%); [11] who found that the microorganisms involved in camel liver at Sharkia were Staphylococcus aureus (18.87%), Streptococcus pyogenes (7.55%), Corynebacterium pyogenes (7.55%), Proteus vulgaris (24.53%), Mycobacterium bovis (7.55%), Clostridium perfringens (15.09%), Closteridium novyi (7.55%) and Candida albicans (11.32%) and [5] who reported that the isolated bacteria in liver of buffaloes at Menoufiea governorate were Staphylococcus aureus (36.51%), E. coli (14.29%), Streptococcus pyogenes (12.70%), Corynebacterium pyogenes (9.52%), Clostridium perfringens (9.52%), Klebsiella pneumoniae (7.94%), Citrobacter freundi (6.35%) and Proteus vulgaris (3.17%).

Corynebacterium ovis was isolated from liver abscesses in our investigated sheep (12%). Similar results were obtained by **[8]** who isolated *Coryne. ovis* from hepatic abscess of sheep in Iran (12.8%) meanwhile, higher incidence was reported by **[21]** who isolated

it from liver abscesses in sheep and goat in Iran (42.86%) and [9] who found C. ovis in liver abscesses of sheep in Iran (42%). In this work, the hepatic affections induced by Staphylococcus aureus infection were almost similar in all the investigated farm animals in the form of multiple abscesses. Microscopically, finely granular eosinophilic and basophilic structureless substances infiltrated with neutrophils and sometimes surrounded with fibrous capsule were detected. These results were in a complete agreement with those fore-mentioned by [17, 22, 8] in sheep, [23, 6] in cattle. Meanwhile, these results were in a partial agreement with [10] who isolated Staph. aureus from cases of abscesses, necrosis and cirrhosis in camel, and disagree with [24] who isolated Staph. aureus from cases of necrosis and cirrhosis in buffaloes. In the present study, the liver lesions due to corynebacterial infection were mostly in the form of gross abscesses oozing greenish inspissated pus on cutting. Microscopically, pus appeared as eosinophilic necrotic material infiltrated with polymorphonuclear leucocytes especially neutrophils and enclosed in fibrous capsule was reported. These results were in a complete compliance with those obtained by [25, 13, 24, 7]. Suppurative hepatitis caused by Corynebacterium pyogenes in cattle was appeared grossly as irregular greenish areas containing greenish viscid pus. Microscopically, multiple foci of necrosis infiltrated with neutrophils, macrophages and lymphocytes in the hepatic tissue were seen. These results were not in compliance with [17] who isolated pseudomonas spp. from lesion of suppurative hepatitis in sheep. In our work one liver of camel infected with Coryne. pyogenes mixed with F. necrophorum showing diffuse hemorrhage and coagulative necrosis in the hepatic parenchyma. Nearly similar findings were found by [12] who noticed coagulative necrosis in liver of diarrheic calves associated to F. necrophorum infection. Moreover, chronic hepatitis due to Corynebacterium pyogenes characterized by hepatic cirrhosis and mononuclear leucocytic infiltration was also seen in our investigated camels. Similar results were mentioned previously by [10]. Our results denoted that *Escherichia coli* was isolated from two types of hepatic lesions; the first type was in the form of variable sized abscesses which oozed creamy white viscid pus occasionally mixed with whitish caseated flakes. Microscopically, central heterophilic necrosis surrounded with zone of inflammatory reaction and fibrosis. These lesions were in a complete agreement with that obtained by [17, 26] in sheep, [27, 6] in cattle, [13, 7] in buffaloes and [10] in camels. Meanwhile, the second type of liver lesion due to E.coli as a causative agent was necrotic hepatitis manifested grossly by multiple pale discrete foci of soft texture scattered on the hepatic surface. Microscopically, areas of coagulative necrosis infiltrated with neutrophils, macrophages and

lymphocytes in camels of our study. These lesions agree with that of **[10]** and disagree with **[11]** who isolated *Clostridium perfringens* and *Protreus vulgaris* from coagulative necrosis of liver of camels.

Final conclusion revealed that the incidence of the causative microorganisms and their lesions differed in farm animals.

REFERENCES

- Blood D. C., Radostitis O. M. and Handerson J. A. (1989): Veterinary Medicine (a textbook of the diseases of cattle, sheep, pigs, goats and horses), 6th Ed.,Bailliere Tindall Cassell Ltd, UK.
- 2. **Kelly W.R. (1985):** The liver and biliary system in: pathology of domestic animals. Jubb K.V.F., Kennedy P.C. And Palmer N., 3rd Ed., Vol. 2, Academic Press, Icn., U.S.A.
- 3. Nagaraja T.G. and Chengappa M.M. (1998): Liver abscesses in feedlot cattle: A review J. Anim. Sci., 76: 287298.
- Lechtenberg K.F., Nagaraja T.G., Leipold H.W. and Chengappa M. M. (1988): Bacteriologic and histologic studies of hepatic abscesses in cattle. Am. J. Vet. Res., 49 (1): 58-62.
- 5. Atwa E.I. and Talkhan O.F.A. (2008): Bacteriological affections of livers and the associating serum biochemical changes in buffaloes in Menoufiea governorate. Zag. Vet. J. 36 (1): 1-10.
- Badr S.I. and Nasr E.M. (2009): Histopathological and bacteriological studies on livers affected with fascioliasis in cattle. Egypt .J.Comp. Pathol. & Clinic. Pathol. 22 (1) 19-45.
- 7. Abo El Fetouh E.H., Kadry M.B. and Alam T.H. (2010): Bacteriological, parasitological and pathological studies on some liver affections of buffaloes in Sharkia governorate. Zag. Vet. J. 38 (4): 61-75.
- 8. Ghadrdan-Mashhadi A. and Yosefi H. (2004): A survey on some bacteriological and pathological aspects of sheep liver. International Society for Animal Hygiene- Saint- Malo- 2004.
- 9. Tehrani A., Javanbakht J., Hassan M. A.M., Zamani M., Rajabian M., Akbari H and Shafei R. (2012): Histopathological and bacteriological study on hepatic abscesses of herrik sheep. J Med Microb Diagn., 1(4).
- 10. **Darwish F.M.M. (1996):** Pathological studies on some liver affection in camels. Ph.D. Thesis, Fac. Vet. Med. Cairo Univ.
- 11. Mohamed M.H., El-Saied T.I. and Attia U.H. (1997): Pathological studies on some liver affections in camels at Sharkia Governorate. Egypt .J.Comp. Pathol & Clinic Pathol. 10 (2): 17-27.
- 12. El-Shahawy H.S. and Abdel-Gaied S.S. (2006): Histopathological studies in young diarrheic calves due to some anaerobic bacteria. J. Egypt. Vet. Med. Assoc., 66(4): 51-66.
- 13. Zaki E. R., Yanny A.A., Shalaby B. and Sobhy N.M. (2000): Bacteria causing liver affection in buffaloes. J. Egypt. Vet. Med. Ass., 60(2): 77-87.

- 14. Boone D.R. and Castenholz R.W. (2001): Bergey's Manual of Systemic Bacteriology. 2nd Ed., Vol. 1, USA.
- 15. Koneman E.W., Allen S.D., Dowell V.R. and Sommers H.M. (1992): Color atlas and textbook of diagnostic microbiology, 4th ed., J.B. Lip. Co., London.
- 16. **Bancroft J.D. and Marilyn A. (2002):** Theory and practice of histological techniques. 5th ed. London, Edinburgh, New York, Philadelphia, St.Louis, Sydney and Toronto.
- 17. El-Mashad A.I., (1987): Morphopathological studies on some liver affection in sheep. M.V.Sci. Theses Fac.Vet. Med. Zagazig university, Benha branch.
- El-Dakhly K. M., Hassan W. H. and Lotfy H. S. (2007): Some parasitic and bacterial causes of liver affections in ruminants. Beni-Suef Vet. Med. J., 5th Sci. Conf., p: 62-68.
- 19. Radostitis O.M., Blood D.C., Gay C.C. and Hinchcliff K.W. (2000): Veterinary medicine 9th ed. W.B. Saunders, China.
- 20. Rosa J.S., Johson E.H., Alves F.S. and Santos L.F. (1989): A retrospective study of hepatic abscesses in goats: Pathological and microbiological findings. Br. Vet. J., 145: 73-76.
- 21. Araghi–Sooreh A. and Firuzi M. (2011): A bacteriologic study on hepatic abscesses of goats slaughtered at the Urmia abattoir. J. Fac. Vet. Med., Islamic Azad Univ., Iran, 5 (1): 73.
- 22. El-Sayed Z.M., El-Seedy F.R., El-Bardisy M.M. and Kamel S.M. (1991): A bacteriological study of sheep liver abscesses. Vet. Med. J., Giza 39(1): 91-103.
- 23. Abd El-Fattah A. M., Saied A. M. and Saied Ash. M. (1995): Bacteriological assessment of inflammed cattle livers with special view to hepatic abscesses.

Assiut Vet. Med. J., 32 (63): 96-101.

- 24. Sayed S.M., Sayed G.M. and El-Nisr N.A. (2008): Clinico-diagnostic studies on hepatic affections of aged buffaloes. Assiut Vet. med. J., 54 (117): 310-328.
- 25. Farag M.M.E (1975): Histopathological studies of necrotic foci in the liver of sheep. M. V. Sc. Thesis Fac. Vet. Med. Cairo Univ.
- 26. Scanlan C. M. and Edwards J. F. (1990): Bacteriologic and pathologic studies of hepatic lesions in sheep. Am. J. Vet. Res., 51(3): 363-366.
- 27. Lim J.S., Kang M., Han D.U., Wee S.H and Chung M.H. (1997): Pathological studies on hepatic lesions of slaughtered Korean beef cattle. Kor. J. Vet. Publ. Hlth., 21(1): 3-10.

دراسات باثولوجيه مقارنة على اصابات الكبد البكتيرية في حيوانات المزرعة

محمد جودة السيد برعى ، عبد الرحيم امين ناجى ،محمود سالم جاب الله، عبدالباسط اسماعيل المشد ، شوقى أحمد جودة السي

قسم الباثولوجيا ، كلية الطب البيطري ، جامعة بنها

الملخص

تم دراسة الاصابات المرضية البكتيرية فى كبد بعض حيوانات المزرعة وتحديد مسبباتها ونسبها المختلفة. وذلك بفحص كبد (١٠٠٠ غنم ، ٨٢٠ بقر ، ١٧٤٠ جاموس ، و ١٣٦٠ جمل، مختلفة السن والجنس) و تجميع المصاب منها ظاهريا فى مجازر القاهرة و الجيزة والقليوبية فى الفترة من يناير 2011 الى مارس ٢٠١٣ لفحصها هستوباثولوجيا وبكتريولوجيا. وقد بلغت نسبة الاصابة البكتيرية فى كبد الغنم ، البقر ، الجاموس، والجمال ٥ %، ٣٠٠٣ %، ١.٢٩ %، و ١٩.١% على التوالى. وقد أوضحت نتيجة الفحص البكتريولوجى أن المسببات البكتيرية الهوائية الاكثر عزلا هى المكور العنقودى الذهبى ، ميكروب الكورينى الصديدى، الميكروب العصوى القولونى بينما عزل ميكروب السل الكاذب من اصابات كبدية فى الغنم فقط. أما أكثر البكتريا الاهوائية عزلا هى الكلستريديم برفرنجنز ، و الفيوز وبكتريوم نكروفورم وتكون مختلطة مع الهوائيات. بالنسبة للتغيرات المرضية للاصابات البكتيرية كانت فى صورة خراريج بينما وجد نخر تجلطى فى الخلايا